

Reference curves for CD4 T-cell count response to combination antiretroviral therapy in HIV-1-infected treatment-naïve patients

V Bouteloup,^{1,2,3} C Sabin,⁴ A Mocroft,⁴ L Gras,⁵ N Pantazis,⁶ V Le Moing,⁷ A d'Arminio Monforte,⁸ M Mary-Krause,⁹ B Roca,¹⁰ JM Miro,¹¹ M Battegay,¹² N Brockmeyer,¹³ J Berenguer,¹⁴ P Morlat,^{2,15} N Obel,¹⁶ S De Wit,¹⁷ G Fätkenheuer,¹⁸ R Zangerle,¹⁹ J Ghosn,^{20,21} S Pérez-Hoyos,^{22,23} M Campbell,²⁴ M Prins,^{25,26} G Chêne,^{1,2,3,27} L Meyer,^{28,29} M Dorrucchi,³⁰ C Torti³¹ and R Thiébaud^{2,3,27} The Standard Reference Distribution of CD4 Response to HAART Project Team for the Collaboration of Observational HIV Epidemiological Research Europe (COHERE) in EuroCoord*

¹CIC 1401, CHU de Bordeaux, Bordeaux, France, ²INSERM U1219 – Centre Inserm Bordeaux Population Health, Université de Bordeaux, Bordeaux, France, ³ISPED, Centre INSERM U1219-Bordeaux Population Health, Université de Bordeaux, Bordeaux, France, ⁴Research Department of Infection & Population Health, UCL, London, UK, ⁵Stichting HIV Monitoring, Amsterdam, The Netherlands, ⁶Department of Hygiene, Epidemiology & Medical Statistics, Athens University Medical School, Athens, Greece, ⁷Montpellier University, Montpellier, France, ⁸Infectious Diseases Unit, Department of Health Sciences, San Paolo University Hospital, Milan, Italy, ⁹INSERM, Institut Pierre Louis d'épidémiologie et de Santé Publique (IPLESP UMRS 1136), UPMC Univ Paris 06, Sorbonne Universités, F-75013, Paris, France, ¹⁰Hospital General of Castellón, Castellón, Spain, ¹¹Infectious Diseases Service. Hospital Clinic – IDIBAPS, University of Barcelona, Barcelona, Spain, ¹²Division of Infectious Diseases and Hospital Epidemiology, Department of Clinical Research, University Hospital of Basel, Basel, Switzerland, ¹³Department of Dermatology, Venerology – Center for Sexual Health and Medicine, Ruhr-Universität Bochum, Bochum, Germany, ¹⁴Instituto de Investigación Sanitaria Gregorio Marañón (IiSGM), Hospital General Universitario Gregorio Marañón, Madrid, Spain, ¹⁵Service de Médecine Interne et Maladies Infectieuses, Hôpital Saint-André, Bordeaux, France, ¹⁶Department of Infectious Diseases, Copenhagen University Hospital, Copenhagen, Denmark, ¹⁷Department of Infectious Diseases, St Pierre University Hospital, Université Libre de Bruxelles, Brussels, Belgium, ¹⁸Department of Internal Medicine, University of Cologne and German Centre for Infection Research (DZIF), Cologne, Germany, ¹⁹Medical University Innsbruck, Innsbruck, Austria, ²⁰APHP, Unité Fonctionnelle de Thérapeutique en Immuno-Infectiologie, Centre Hospitalier Universitaire Hôtel Dieu, Paris, France, ²¹Faculté de Médecine Site Necker, Sorbonne Paris Cité, Université Paris Descartes, EA 7327, Paris, France, ²²Vall d'Hebrón Institut de Recerca (VHIR), Barcelona, Spain, ²³Universitat Autònoma de Barcelona, Barcelona, Spain, ²⁴CHIP, Department of Infectious Diseases, Rigshospitalet, University of Copenhagen, Copenhagen, Denmark, ²⁵Division of Infectious Diseases, Department of Internal Medicine, Center for Infection and Immunity Amsterdam (CINIMA), Academic Medical Center, Amsterdam, The Netherlands, ²⁶Department of Infectious Diseases, Public Health Service, Amsterdam, The Netherlands, ²⁷CHU de Bordeaux, Pole de Sante Publique, Service d'Information Medicale, F-33000, Bordeaux, France, ²⁸INSERM, U1018, Epidemiology of HIV, Reproduction, Paediatrics, CESP; University Paris-Sud, Paris, France, ²⁹Department of Public Health and Epidemiology, Bicêtre Hospital, AP-HP, Le Kremlin Bicêtre, Paris, France, ³⁰Department of Infectious, Parasitic and Immune-mediated Diseases, Istituto Superiore di Sanità, Rome, Italy and ³¹Unit of Infectious and Tropical Diseases, Department of Medical and Surgical Sciences, University “Magna Graecia”, Catanzaro, Italy

Correspondence: Pr Rodolphe Thiébaud, Centre Inserm U1219, Université Bordeaux, 146 rue Léo Saignat, Case 11, 33076 Bordeaux Cedex, France. Tel: +33557571393; fax: +33557571578; e-mail: rodolphe.thiebaud@isped.u-bordeaux2.fr

and

Vincent Bouteloup, CIC 1401, CHU de Bordeaux, ISPED, 146 rue Léo Saignat, Case 11, 33076 Bordeaux Cedex, France. Tel: +33557571393; fax: +33557575713; e-mail: vincent.bouteloup@isped.u-bordeaux2.fr

*See Appendix for a list of the Standard Reference Distribution of CD4 Response to HAART Project Team for the Collaboration of Observational HIV Epidemiological Research in Europe (COHERE) in EuroCoord, who are the authors of this paper, and their affiliations.

Objectives

The aim of this work was to provide a reference for the CD4 T-cell count response in the early months after the initiation of combination antiretroviral therapy (cART) in HIV-1-infected patients.

Methods

All patients in the Collaboration of Observational HIV Epidemiological Research Europe (COHERE) cohort who were aged ≥ 18 years and started cART for the first time between 1 January 2005 and 1 January 2010 and who had at least one available measurement of CD4 count and a viral load ≤ 50 HIV-1 RNA copies/mL at 6 months (± 3 months) after cART initiation were included in the study. Unadjusted and adjusted reference curves and predictions were obtained using quantile regressions.

Results

A total of 28 992 patients were included in the study. The median CD4 T-cell count at treatment initiation was 249 [interquartile range (IQR) 150, 336] cells/ μ L. The median observed CD4 counts at 6, 9 and 12 months were 382 (IQR 256, 515), 402 (IQR 274, 543) and 420 (IQR 293, 565) cells/ μ L. The two main factors explaining the variation of CD4 count at 6 months were AIDS stage and CD4 count at cART initiation. A CD4 count increase of ≥ 100 cells/mL is generally required in order that patients stay 'on track' (i.e. with a CD4 count at the same percentile as when they started), with slightly higher gains required for those starting with CD4 counts in the higher percentiles. Individual predictions adjusted for factors influencing CD4 count were more precise.

Conclusions

Reference curves aid the evaluation of the immune response early after antiretroviral therapy initiation that leads to viral control.

Keywords: antiretroviral treatment monitoring, CD4 response, HIV monitoring, longitudinal data.

Accepted 28 January 2016

Introduction

The improvement of antiretroviral treatments since the mid-1990s has led to a large decrease in the incidence of severe morbidities [1] in those living with HIV, with a substantial increase in life expectancy as a result [2]. For an individual starting combination antiretroviral therapy (cART) for the first time, the current goal of treatment is to ensure that the individual's viral load reaches undetectable levels as soon as possible after initiation of cART with his/her CD4 T-cell count increasing soon thereafter [3]. Hence, a good virological response to cART is usually defined as an undetectable viral load within the first 6 months after cART initiation. A good immunological response, however, is less clearly defined. CD4 counts in the HIV-negative population usually lie within the range of 500–1500 cells/ μ L [4]. Among those with HIV infection, a higher CD4 T-cell count is associated with a lower risk of clinical progression [5,6]; in particular, the longer an HIV-positive individual is able to maintain a CD4 count > 500 cells/ μ L, the closer is his/her life expectancy to that of the general population [7]. The risk of clinical progression appears to be more strongly associated with the absolute CD4 count at a given time rather than the rate of increase in the CD4 count [8,9].

Despite the difficulty of defining a good immune responder, it is important that clinicians are provided with a reference so that they are able to evaluate an individual's CD4 T-cell response in the early months after starting cART. This will allow them to make decisions about intervening in the management of the patient; for instance, decisions about the appropriate frequency of clinical monitoring. Our aim was to provide 'reference curves' for CD4 T-cell responses during the first 12 months of cART for patients with virological suppression, according to the characteristics of the patients at cART initiation. It is hoped that these curves will allow clinicians to determine how an individual's CD4 count response compares to those of other patients who started the same type of cART under the same conditions.

Methods

Patients

The Collaboration of Observational HIV Epidemiological Research Europe (COHERE) is a collaboration involving 39 cohorts from across Europe and is part of the EuroCoord network (www.EuroCoord.net). COHERE was

established in 2005 with the aim of conducting epidemiological research on prognosis and outcome in HIV-positive persons, which the individual contributing cohort studies cannot address themselves because of small sample sizes or heterogeneity of specific subgroups of HIV-positive persons. Local ethical committee and/or other regulatory approval was obtained as applicable according to local and/or national regulations in all participating cohorts. Each cohort study group submits data using the standardized HIV Collaboration Data Exchange Protocol (HICDEP) [10], including information on patient demographics, use of cART, CD4 counts, AIDS and deaths. Further details can be found at http://www.eurocoord.net/partners/founding_networks/cohere.aspx. Data were pooled in September 2011 within COHERE in EuroCoord (www.cohere.org and www.EuroCoord.net). Data on 27 cohorts across 35 European countries were provided for the present analysis. All persons aged ≥ 18 years who started cART for the first time between 1 January 2005 and 1 January 2010 and who had at least one available measurement of CD4 count (in cells/ μ L) and a viral load ≤ 50 HIV-1 RNA copies/mL 6 months (± 3 months) after cART initiation were included in the study. Patients were considered to have maintained viral suppression at month 6 when at least one HIV-1 RNA measurement was available within 6 months after cART initiation (including the measurement at month 6) and the HIV-1 RNA measurement at month 6 was < 50 copies/mL (previous measurements could be > 50 copies/mL). cART was defined as any regimen that contained at least three drugs, including a protease inhibitor (PI), a nonnucleoside reverse transcriptase inhibitor (NNRTI), an entry inhibitor or an integrase inhibitor, or that contained three nucleoside reverse transcriptase inhibitors (NRTIs), of which one was abacavir. Baseline was defined as the date of cART initiation. The follow-up was censored first at 9 months (i.e. 6 + 3 months) and then at 15 months (i.e. 12 + 3 months) because we were interested in the short-term response of CD4 count.

Strategy for statistical analyses

Any estimate of the effect of a given factor on CD4 T-cell dynamics is likely to be significant in this large study because of the statistical power conferred by the size of the data set. Therefore, factors used for stratified analyses or included in the regression model were defined *a priori*. Baseline factors known to be associated with CD4 T-cell change after cART initiation selected for the analyses were: CD4 T-cell count, AIDS clinical stage, age, HIV transmission group [especially injecting drug use (IDU)], sex, presence *vs.* absence of hepatitis C virus (HCV) coinfection,

Table 1 Baseline characteristics

Characteristic	Analysed population (<i>n</i> = 28 992)
Gender, male [<i>n</i> (%)]	21 130 (72.9)
Age at cART initiation (years) [median (IQR)]	39 (33, 46)
Duration of follow-up after initiating cART (years) [median (IQR)]	2.3 (1.3, 3.5)
HIV transmission risk group [<i>n</i> (%)]	
Heterosexual	11 601 (40.0)
Men who have sex with men	12 547 (43.3)
Injecting drug use	1952 (6.7)
Other/unknown	2892 (10.0)
CDC stage C [<i>n</i> (%)]	
No	21 626 (74.6)
Yes	6144 (21.2)
Unknown	1222 (4.2)
HCV positive [<i>n</i> (%)]	2250 (7.8)
HIV-1 RNA [<i>n</i> (%)]	
< 50 copies/mL	2738 (9.4)
50–9999 copies/mL	4346 (15.0)
10 000–99 999 copies/mL	9730 (33.6)
$\geq 100 000$ copies/mL	8731 (30.1)
Unknown	3447 (11.9)
cART initiation year [<i>n</i> (%)]	
2005	5342 (18.4)
2006	5864 (20.2)
2007	6266 (21.6)
2008	6958 (24.0)
2009	4562 (15.7)
Number of CD4 measurements within 9 months [median (IQR)]	3 (2, 4)
CD4 count prior to cART initiation (cells/ μ L) [median (IQR)]	249 (150, 336)
0–199 cells/ μ L [<i>n</i> (%)]	9647 (35.6)
200–349 cells/ μ L [<i>n</i> (%)]	11 395 (42.1)
350–499 cells/ μ L [<i>n</i> (%)]	3769 (13.9)
≥ 500 cells/ μ L [<i>n</i> (%)]	2286 (8.4)
Time from baseline to 6-month HIV-1 RNA measurement (months) [median (IQR)]	5.9 (5.1, 6.7)
Duration of first-line regimen (months) [median (IQR)]	13.5 (5.5, 26.5)
First-line cART regimen [<i>n</i> (%)]	
PI/r-based	12 706 (43.8)
NNRTI-based	14 665 (50.6)
PI- and NNRTI-based	328 (1.1)
Unboosted PI-based	607 (2.1)
Other*	686 (2.4)

cART, combination antiretroviral therapy; IQR, interquartile range; CDC, Centers for Disease Control and Prevention; HCV, hepatitis C virus; PI, protease inhibitor; NNRTI, nonnucleoside reverse transcriptase inhibitor. *Regimens containing abacavir (*n* = 496), integrase inhibitors (*n* = 151), fusion inhibitors (*n* = 24) and other combinations (*n* = 15).

HIV-1 RNA level, year of cART initiation and type of cART. cART regimen was classified as: ritonavir-boosted PI-based, NNRTI-based, both PI- and NNRTI-based, unboosted PI-based, or another regimen that did not include either a PI or an NNRTI. Integrase or fusion inhibitor-based regimens were too infrequently used to constitute specific subgroups and were included in the previously described categories. The distribution of CD4 T-cell count at 6 (12) months was then described by using the closest measurement between 3 (9) and 9 (15) months.

Quantile regression

Quantile regression was performed on repeated measurements of CD4 count available for each individual until 9 months and then until 15 months. Quantiles of specific interest were the 5, 10, 25, 50th (i.e. median), 75, 90 and 95th percentiles. The models were adjusted for time with squared and cubic effects to allow enough flexibility to fit a nonlinear evolution of CD4 count over time. A set of models (one for each percentile of interest) was fitted without including any other covariates in the models after stratifying according to the baseline CD4 count (0–199, 200–349, 350–499 and ≥ 500 cells/ μ L). These models were used to draw reference curves for the overall distribution of CD4 T-cell responses among individuals in the study. A second set of models was then fitted after additionally including adjustment for the covariates listed above; this set of models can be used to make predictions for individual responses where individual characteristics are taken into account. The models presented were fitted using PROC QUANTREG in SAS v9.3 (SAS Institute, Cary, NC, USA). Robustness analyses with median regression including a random intercept to take into account

correlation of repeated measurements in patients yielded similar results [11], as did a nonparametric approach [12].

Reference curves can be generated for each individual using a web tool developed with SHINY, a web application framework for R (<http://shiny.rstudio.com/>).

Results

Study population

A total of 28 992 patients were included in the study. A description of the study population is provided in Table 1 (Fig. 1). Compared with included individuals (Table S1), those who were excluded were younger (median 38 *vs.* 39 years for included individuals), were less frequently male (69 *vs.* 73%, respectively), were less frequently men who have sex with men (MSM) (35 *vs.* 43%, respectively), and had more frequently started a PI/r-based cART regimen (51 *vs.* 44%, respectively) ($P < 10^{-4}$ for all comparisons). The difference in baseline CD4 count at cART initiation was marginal (Wilcoxon rank sum test $P = 0.066$). The median number of CD4 T-cell

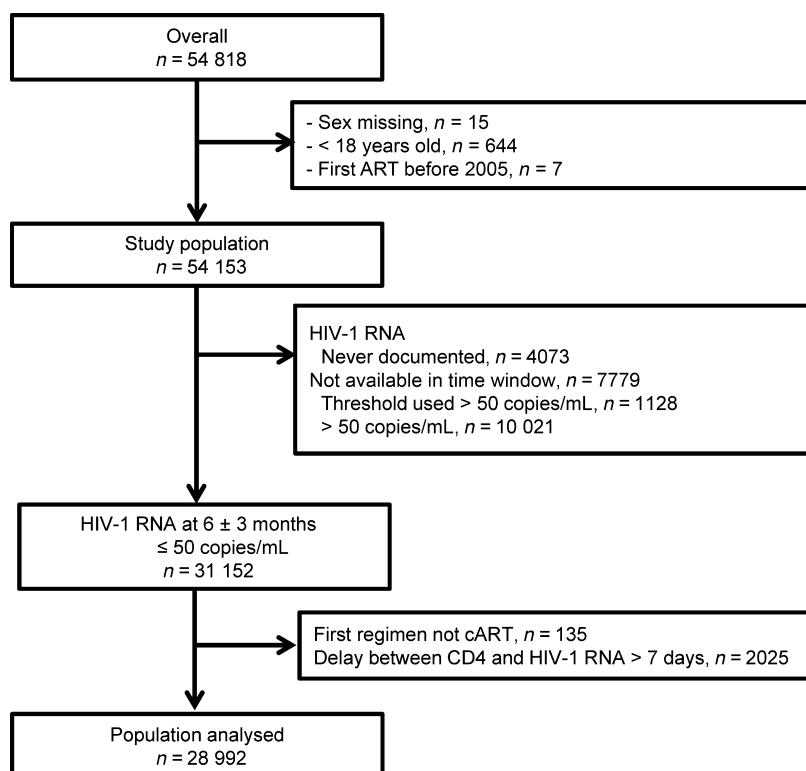


Fig. 1 Flow chart of patient selection. ART, antiretroviral therapy; cART, combination antiretroviral therapy.

measurements available per patient was 3 [interquartile range (IQR) 2, 4].

Global overview of the quantiles of CD4 T-cell changes

The median (IQR) observed CD4 count at treatment initiation was 249 (150, 336) cells/ μL (Figs 2 and 3). The median (IQR) observed CD4 counts at 6, 9 and 12 months were 382 (256, 515), 402 (274, 543) and 420 (293, 565), respectively. Box plots of the CD4 T-cell response at month 6 according to various baseline characteristics are presented in Fig. 2. The two main factors explaining the variation of CD4 count at 6 months were AIDS stage at cART initiation and, as expected, the baseline CD4 count. A similar picture was seen at 12 months (Fig. S1).

Figure 3 shows the predicted percentiles of CD4 count after cART initiation for the whole population. The figure shows that a CD4 count increase of at least 100 cells/ mL is generally required in order that patients stay 'on track' (i.e. at the same percentile as when they started), with slightly higher gains required to stay on track for those starting with CD4 counts in the higher percentiles (Table S2). For example, the median line demonstrates

that patients who started cART at the median level of 251 cells/ μL needed to have a CD4 count of 367 cells/ μL at 6 months after cART initiation to remain on the median line. The gain required to stay on track was somewhat larger for those initiating cART at higher percentiles of the CD4 T-cell distribution. For instance, a patient starting cART with a CD4 count of 500 cells/ μL (roughly equating to the 90th percentile) would need to have attained a CD4 count of 650 cells/ μL by 6 months in order to remain on the same percentile line.

As examples, the trajectories of two specific patients are depicted in Fig. 3. The first one (purple line) represents an individual who was female and 30 years old who started cART (lopinavir/r + zidovudine/lamivudine) with a CD4 count of 390 cells/ μL , representing the 82th percentile of baseline values. Her CD4 count had increased to only 438 cells/ μL over the first 7.9 months. This increase was modest, but was more obviously seen to be a poor response when looked at relative to the population as a whole, representing a drop from the 82th percentile to the 61th percentile. In the second example (green line), cART (efavirenz + rilpivirine + tenofovir/emtricitabine) was started at 112 cells/ μL (the 18th percentile); by 7.3 months, this individual's CD4 count had

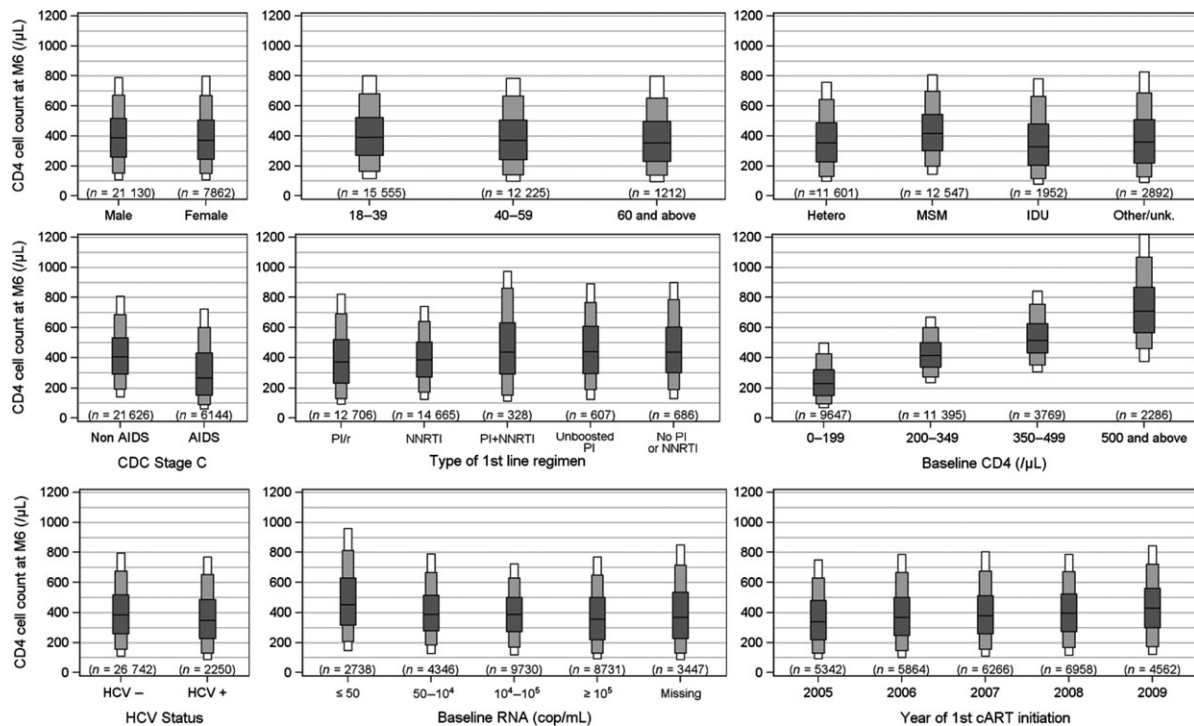


Fig. 2 Box plot of CD4 count at month 6 according to various baseline characteristics. Boxplots show, respectively, the 5, 10, 25, 50, 75, 90 and 95th percentiles. M6, month 6. cART, combination antiretroviral therapy; CDC, Centers for Disease Control and Prevention; HCV, hepatitis C virus; IDU, injecting drug use; MSM, men who have sex with men; NNRTI, nonnucleoside reverse transcriptase inhibitor; PI, protease inhibitor; r, ritonavir.

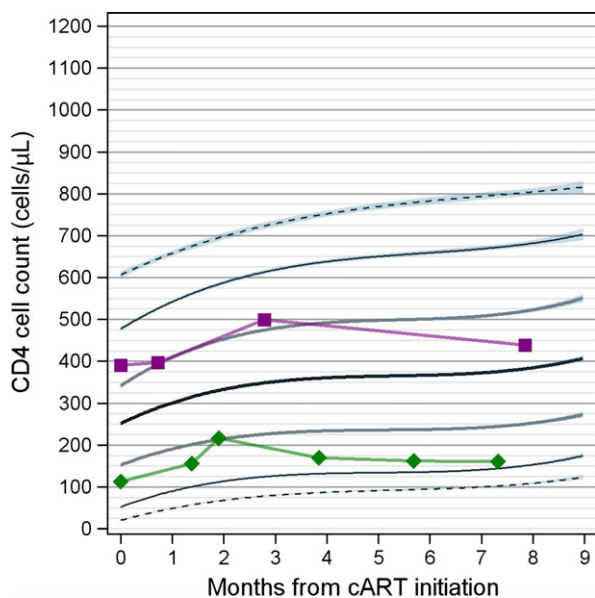


Fig. 3 Percentiles (5, 10, 25, 50, 75, 90 and 95th) of CD4 count over time from combination antiretroviral therapy (cART) initiation. Two individual trajectories are added for illustration: one (purple line) drops from the 82th percentile to the 61th percentile, and the other (green line) starts at the 18th percentile and has dropped to the 12th percentile at 7 months. cART, combination antiretroviral therapy.

reached 161 cells/ μL , representing the 12th percentile, which could be considered a bad response. This figure illustrates how this type of representation could help clinicians when evaluating the CD4 count change after cART initiation.

Factors associated with CD4 count

Factors associated with CD4 count were evaluated for each percentile of interest using quantile regression (Table 2). For instance, the 25th percentile for women was 14 cells/ μL higher than that for men and this difference was consistent over the other percentiles (14 and 11 cells/ μL for the 50 and 75th percentiles, respectively). However, the impact of some baseline characteristics varied according to the percentile. AIDS clinical stage had the greatest impact for individuals with CD4 counts around the 25th (-111 cells/ μL) and 50th (-115 cells/ μL) percentiles, whereas the type of cART regimen had the greatest impact for those with counts in the highest (≥ 75 th) percentiles. For instance, those starting a regimen including both a PI and an NNRTI had CD4 counts 103 cells/ μL higher at 6 months than patients who started a regimen including only boosted PI in addition to NRTIs.

Our predictions were then further refined using the measured baseline characteristics in a multivariable quantile regression. Examples are provided in Fig. 4 for the two patients whose data are presented in Fig. 3. The first patient, who started with a CD4 count of 390 cells/ μL at the 82th percentile (solid purple line, left part), is classified as starting at the 44th percentile after adjustment for sex, age, transmission risk group, clinical stage, HIV RNA and cART regimen at baseline (dotted purple line, left part). She had attained a CD4 count of 438 cells/ μL at 7.9 months after cART initiation, which corresponds to the 61th percentile in the crude analysis (solid purple line, right part). However, taking the individual's characteristics into account, the patient's attained CD4 count is found to lie on only the 22th percentile after adjustment (dotted purple line, right part). Therefore, this increase of only 48 cells/ μL means that, in relative terms, the patient has switched from the 44th percentile to the 22nd percentile; this is clearly a poor response. The second patient (solid and dotted green lines) presented with a not so bad response, as he started at the 62th adjusted percentile with a CD4 count of 112 cells/ μL and dropped to the 30th adjusted percentile with a CD4 count of 161 cells/ μL at 7 months, as compared to the crude percentiles which were 18th at baseline and 12th at 7 months.

Discussion

The CD4 T-cell reference curves presented here provide an indication of how the immunological response of an individual patient may compare to that of a large sample of HIV-infected patients who have achieved viral control with potent antiretroviral therapy. These reference curves can easily be drawn using the online tool we developed, available at <http://shiny.isped.u-bordeaux.fr/CD4refcurves/>.

The definition of immune nonresponse is highly variable in the literature [13–16]. One obvious explanation for the variability is that the latest absolute CD4 count is more closely associated with clinical prognosis than the slope of the CD4 T-cell increase [9]. Hence, there is no obvious threshold for a *good* immune response other than having the highest possible CD4 count. This is why we propose that individual trajectories are referred to the distribution of the whole population, rather than considering whether the absolute count is above or below an arbitrary threshold value.

Using our reference curve, an immune nonresponder may be defined as a person who, despite cART, was not able to maintain an immune response that allowed her/him to remain on the same percentile curve as at the start of cART. In the figures, we provide quartiles and extreme percentiles. Although there is no evidence for an additional

Table 2 Factors associated with the CD4 count according to percentile (25, 50 and 75th percentiles): multivariate quantile regressions of CD4 count

	25th percentile		50th percentile		75th percentile	
	Beta (95% CI)	P	Beta (95% CI)	P	Beta (95% CI)	P
Female (reference: male)	13 (11, 16)	< 10 ⁻⁴	13 (11, 16)	< 10 ⁻⁴	10 (6, 13)	< 10 ⁻⁴
Age (per 10-year increase)	-8 (-9, -7)		-9 (-10, -8)		-9 (-10, -8)	< 10 ⁻⁴
HIV transmission risk group (reference: MSM)						
Heterosexual	-59 (-61, -57)	< 10 ⁻⁴	-60 (-62, -57)	< 10 ⁻⁴	-62 (-65, -58)	< 10 ⁻⁴
Injecting drug use	-67 (-72, -62)		-66 (-71, -61)		-55 (-62, -48)	
Other/unknown	-64 (-67, -60)		-61 (-65, -57)		-54 (-60, -49)	
CDC stage C (reference: A/B)						
Yes	-111 (-113, -109)	< 10 ⁻⁴	-115 (-117, -112)	< 10 ⁻⁴	-89 (-93, -86)	< 10 ⁻⁴
Unknown	1 (-4, 6)		9 (4, 14)		11 (5, 18)	
HCV coinfection (reference: no)	-6 (-10, -3)	7.10 ⁻⁴	-6 (-10, -3)	4.10 ⁻⁴	-12 (-17, -7)	< 10 ⁻⁴
HIV-1 RNA (reference: < 50 copies/mL)						
50–9999 copies/mL	-32 (-36, -28)	< 10 ⁻⁴	-62 (-67, -58)	< 10 ⁻⁴	-105 (-111, -98)	< 10 ⁻⁴
10 000–99 999 copies/mL	-42 (-46, -38)		-72 (-76, -68)		-122 (-128, -116)	
≥ 100 000 copies/mL	-69 (-72, -65)		-91 (-96, -87)		-126 (-132, -120)	
Unknown	-62 (-67, -58)		-79 (-84, -74)		-100 (-107, -92)	
First-line cART regimen (reference: PI/r-based)						
NNRTI-based	8 (6, 10)	< 10 ⁻⁴	-2 (-5, -0)	< 10 ⁻⁴	-25 (-28, -22)	< 10 ⁻⁴
PI- and NNRTI-based	40 (31, 50)		57 (47, 67)		103 (85, 121)	
Unboosted PI-based	32 (25, 40)		48 (39, 56)		56 (46, 66)	
Other	52 (45, 59)		51 (43, 59)		49 (39, 58)	
Year of cART initiation (reference: 2009)						
2005	-48 (-51, -45)	< 10 ⁻⁴	-65 (-68, -62)	< 10 ⁻⁴	-70 (-75, -66)	< 10 ⁻⁴
2006	-36 (-39, -32)		-44 (-47, -41)		-45 (-50, -40)	
2007	-26 (-29, -23)		-38 (-41, -35)		-40 (-45, -36)	
2008	-21 (-25, -18)		-29 (-32, -25)		-31 (-36, -27)	

cART, combination antiretroviral therapy; CDC, Centers for Disease Control and Prevention; CI, confidence interval; HCV, hepatitis C virus; PI, protease inhibitor; NNRTI, nonnucleoside reverse transcriptase inhibitor; MSM, men who have sex with men.

value of immune interventions [17] or changing cART regimens in such a situation, research on boosting the immune response is ongoing [18]. In addition, as long as the observed increase for a given patient means that s/he remains on the same percentile and provided that viral replication is controlled, there may be an argument for scheduling fewer visits in the subsequent months. With the availability of more potent antiretroviral drugs with fewer side effects, and a possible overall benefit of starting antiretroviral therapy as early as possible [19], patients will tend to start cART at higher CD4 cell counts. Thus, a patient starting at 200 cells/ μ L could be at the 30th percentile in 2004 but at the 20th percentile in 2014, whereas attainment of a count of 300 cells/ μ L at 6 months may place him/her on the 30th percentile in 2004 but only the 20th in 2014. Therefore, these reference curves need to be regularly updated to remain relevant.

We restricted the analysis to the first 9 months after cART initiation to retain homogeneity among the treatment regimens that were being used (physicians are unlikely to change regimens in the first few months unless there is toxicity) [20]. Furthermore, the increase in CD4 count over the first few months is more pronounced than at later times, partly as a result of the redistribution of T

cells [21]. However, it is likely that adherence is at its highest in the first 9 months, and thus our reference curves are likely to identify those with suboptimal CD4 responses despite relatively good adherence to therapy. The extension of our findings to longer durations of cART exposure is under consideration. The results presented in this work were obtained from statistical analyses that did not account for the correlation of repeated measures performed for each patient. Although the method proposed by Geraci and Bottai [11] (LQMM R package) based on random effects yielded similar results for the median, we found inconsistent results for the other percentiles. However, we assume that the use of a correlation matrix would have a very slight impact on our estimations, considering the high number of patients compared with the limited number of observations for each patient.

Despite these limitations, this work has demonstrated the advantages of having a large sample size with a good representation of the HIV-infected population; this may be difficult to achieve in a single study/cohort. For instance, CD4 T-cell dynamics depend on the baseline level. In those starting cART with low CD4 counts, full immune restoration might be expected to be slow because of the profound immune suppression that has led to

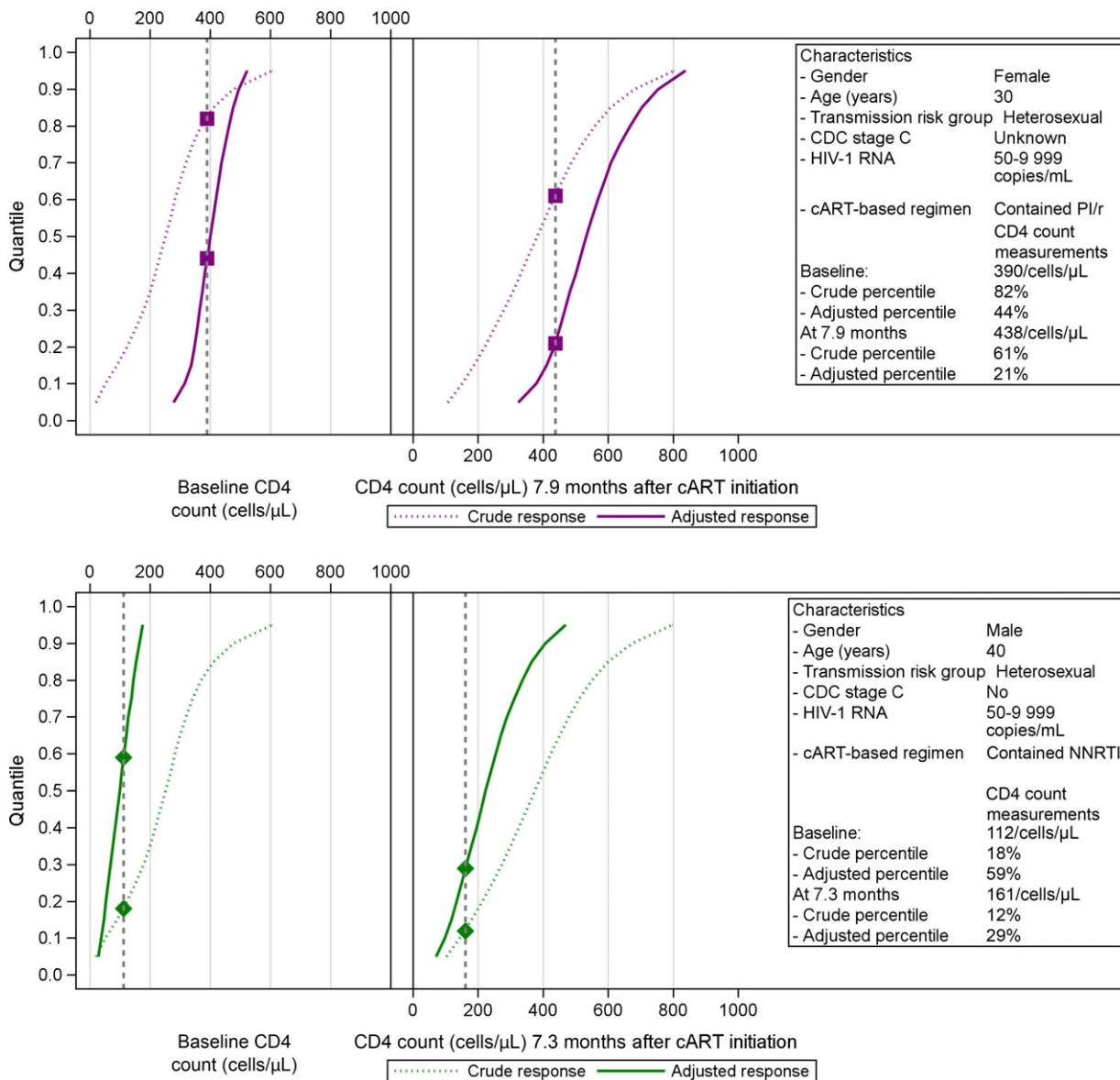


Fig. 4 Example of individual CD4 cell count responses (crude and adjusted) for two patients, data for whom are also presented in Fig. 3. Crude (dotted lines) and adjusted (solid lines) percentiles at baseline (left) and during follow-up (right) are provided according to the measured CD4 count (dashed line). cART, combination antiretroviral therapy; CDC, Centers for Disease Control and Prevention; NNRTI, nonnucleoside reverse transcriptase inhibitor; PI, protease inhibitor; r, ritonavir.

tissue damage in the thymus and lymph nodes [22,23]. However, immune responses are also regulated by homeostatic mechanisms that might result in a ceiling effect, such that those initiating cART with higher CD4 counts might experience less pronounced responses than those starting with lower counts [24]. Hence, looking at the literature, one can find studies where the CD4 T-cell increase was more pronounced in those with higher baseline values [9,25–28], whereas in other studies the CD4 T-cell increase was larger in those starting cART

with lower CD4 T-cell values [29–32]. These differences could be attributable to the differences between study populations and thus indicate a need for large data sets supported by collaborations such as COHERE to derive reference curves.

In conclusion, we propose reference curves for the CD4 count that may be used as an additional tool for the clinician when evaluating responses to cART. A web tool is available at http://shiny.isped.u-bordeaux.fr/CD4_refcurves/.

Acknowledgements

Conflicts of interest: Jose M. Miro has received consulting honoraria from Abbvie, Bristol-Myers Squibb, Gilead Sciences, Merck and Novartis y Sanofi, research and academic grants from Cubist, Gilead, ViiV, Novartis, Merck, Fondo de Investigaciones Sanitarias (FIS) del Instituto de Salud Carlos III (Madrid), Fundación para la Investigación y Prevención del Sida en España (FIPSE; Madrid), Ministerio de Sanidad, Servicios Sociales e Igualdad (MSSSI; Madrid), the National Institutes of Health (NIH; Bethesda, MA, USA) and NEAT, and honoraria for lectures from Abbvie, Bristol-Myers Squibb, Gilead Sciences, Merck, Novartis, ViiV Healthcare and Instituto de Salud Carlos III, Ministerio de Economía y Competitividad, Madrid (Spain) provided funding to Jose M. Miro under a personal intensification research grant # INT15/00168 during 2016. Murielle Mary-Krause received consulting fees from ViiV Healthcare in 2015. Norbert H. Brockmeyer discloses personal compensation or of the organization/institution to which he belongs for activities with the following companies: Bristol-Myers-Squibb, Gilead, ViiV Healthcare, MSD Sharp & Dohme, Janssen-Cilag, Sanofi Pasteur, Hexal, Boehringer Ingelheim, Hologic, Roche, AbbVie and Cepheid. The remaining authors have no conflicts of interest to declare.

Author contributions: The principal contributions made by the authors were as follows. Study design and statistical analysis: Rodolphe Thiébaud and Vincent Bouteloup. Interpretation of results: all authors. Reading and approval of the manuscript: all authors. Drafting of the manuscript: Rodolphe Thiébaud and Vincent Bouteloup. Development of the SHINY web tool: Robin Genuer.

Financial disclosure: The COHERE study group has received unrestricted funding from: the Agence Nationale de Recherches sur le SIDA et les Hépatites Virales (ANRS), France; the HIV Monitoring Foundation, The Netherlands; and the Augustinus Foundation, Denmark. The research leading to these results has received funding from the European Union Seventh Framework Programme (FP7/2007-2013) under EuroCoord grant agreement n° 260694. A list of the funders of the participating cohorts can be found at www.COHERE.org.

Appendix

The Collaboration of Observational HIV Epidemiological Research Europe (COHERE) Group

Steering Committee (contributing cohorts): Robert Zangerle (AHIVCOS), Giota Touloumi (AMACS), Josiane

Warszawski (ANRS CO1 EPF/ANRS CO11 OBSERVATOIRE EPF), Laurence Meyer (ANRS CO2 SEROCO), François Dabis (ANRS CO3 AQUITAINE), Murielle Mary Krause (FHDH-ANRS CO4), Jade Ghosn (ANRS CO6 PRIMO), Catherine Lepout (ANRS CO8 COPILOTE), Linda Wittkop (ANRS CO13 HEPAVIH), Peter Reiss (ATHENA), Ferdinand Wit (ATHENA), Maria Prins (CASCADE), Heiner Bucher (CASCADE), Caroline Sabin (CHIC), Diana Gibb (CHIPS), Gerd Fätkenheuer (Cologne-Bonn), Julia Del Amo (CoRIS), Niels Obel (Danish HIV Cohort), Claire Thorne (ECS), Amanda Mocroft (EuroSIDA), Ole Kirk (EuroSIDA), Christoph Stephan (Frankfurt), Santiago Pérez-Hoyos (GEMES-Haemo), Osamah Hamouda (German ClinSurv), Barbara Bartmeyer (German ClinSurv), Nikoloz Chkhartishvili (Georgian National HIV/AIDS), Antoni Noguera-Julian (CORISPE-cat), Andrea Antinori (ICC), Antonella d'Arminio Monforte (ICONA), Norbert Brockmeyer (KOMPNET), Luis Prieto (Madrid PMTCT Cohort), Pablo Rojo Conejo (CORISPES-Madrid), Antoni Soriano-Arandes (NENEXP), Manuel Battegay (SHCS), Andri Rauch (SHCS), Cristina Mussini (Modena Cohort), Pat Tookey (NSHPC), Jordi Casabona (PISCIS), Jose M. Miro (PISCIS), Antonella Castagna (San Raffaele), Deborah Konopnick (St. Pierre Cohort), Tessa Goetghebuer (St Pierre Paediatric Cohort), Anders Sönnberg (Swedish InfCare), Carlo Torti (Italian Master Cohort), Ramon Teira (VACH), Myriam Garrido (VACH) and David Haerry (European AIDS Treatment Group).

Executive Committee: Stéphane de Wit (Chair; St Pierre University Hospital), Jose M. Miro (PISCIS), Dominique Costagliola (FHDH-ANRS CO4), Antonella d'Arminio-Monforte (ICONA), Antonella Castagna (San Raffaele), Julia del Amo (CoRIS), Amanda Mocroft (EuroSIDA), Dorte Raben (Head, Copenhagen Regional Coordinating Centre) and Geneviève Chêne (Head, Bordeaux Regional Coordinating Centre). Paediatric Cohort Representatives: Ali Judd and Pablo Rojo Conejo.

Regional Coordinating Centres: Bordeaux RCC: Diana Barger, Céline Colin, Christine Schwimmer, Monique Termote and Linda Wittkop; Copenhagen RCC: Maria Campbell, Nina Friis-Møller, Jesper Kjaer, Dorte Raben and Rikke Salbøl Brandt.

Project Leads and Statisticians: Juan Berenguer, Julia Bohlius, Vincent Bouteloup, Heiner Bucher, Alessandro Cozzi-Lepri, François Dabis, Antonella d'Arminio Monforte, Mary-Anne Davies, Julia del Amo, Maria Dorrucchi, David Dunn, Matthias Egger, Hansjakob Furrer, Marguerite Guiguet, Sophie Grabar, Ali Judd, Ole Kirk, Olivier Lambotte, Valériane Leroy, Sara Lodi, Sophie Matheron, Laurence Meyer, Jose M. Miro, Amanda Mocroft, Susana Monge, Fumiyo Nakagawa, Roger Paredes, Andrew Phillips, Massimo Puoti, Michael Schomaker, Colette Smit,

Jonathan Sterne, Rodolphe Thiebaut, Claire Thorne, Carlo Torti, Marc van der Valk, Linda Wittkop and Natasha Wyss.

Standard Reference Distribution of CD4 Response to HAART Project Team (the authors of this paper and their affiliations)

Vincent Bouteloup (Statistician; CIC 1401 Bordeaux University Hospital, ISPED, France), Caroline Sabin (CHIC, University College London, London, UK), Amanda Mocroft (EuroSIDA, Department of Infection and Population Health, University College London, London, UK), Luuk Gras (ATHENA, Stichting HIV Monitoring, Amsterdam, the Netherlands), Nikos Pantazis (AMACS, Department of Hygiene, Epidemiology & Medical Statistics, Athens University Medical School, Athens, Greece), Vincent Le Moing (APROCO-COPILOTE, Montpellier University, Montpellier, France), Antonella d'Arminio Monforte (ICONA, Infectious Diseases, University of Milan, Milan, Italy), Murielle Mary-Krause (FHDH-ANRS C04, Sorbonne Universités, UPMC Univ Paris 06, INSERM, Institut Pierre Louis d'épidémiologie et de Santé Publique (IPLESP UMRS 1136), F75013, Paris, France), Bernardino Roca (VACH, Hospital General of Castellon, Castellon, Spain), Jose M. Miro (PISCIS, Hospital Clinic Universitari, Barcelona, Spain), Manuel Battegay (SHCS, Division of Infectious Diseases and Hospital Epidemiology, Department of Clinical Research, University Hospital of Basel, Basel, Switzerland), Norbert Brockmeyer (KOMNET, Department of Dermatology, Venerology - Center for sexual health and medicine, Ruhr-Universität Bochum, Bochum, Germany), Juan Berenguer [CoRIS, Hospital General Universitario Gregorio Marañón. Instituto de Investigación Sanitaria Gregorio Marañón (IiSGM), Madrid, Spain], Philippe Morlat (AQUITAINE, Univ. Bordeaux, ISPED, Centre INSERM U1219-Epidemiologie-Biostatistiques, Bordeaux, France and Service de Médecine Interne et Maladies Infectieuses, Hôpital Saint-André, Bordeaux, France), Niels Obel (Danish HIV Cohort, Department of Infectious Diseases, Copenhagen University Hospital, Copenhagen, Denmark), Stéphane De Wit (St Pierre Cohort, The Brussels Saint Pierre Cohort, Brussels, Belgium), Gerd Fätkenheuer [Cologne-Bonn, Department of Internal Medicine, University of Cologne and German Centre for Infection Research (DZIF), Cologne, Germany], Robert Zangerle (AHIVCOS, Medical University Innsbruck, Innsbruck, Austria), Jade Ghosn (PRIMO, AHPH, Unité Fonctionnelle de Thérapeutique en Immuno-Infectiologie, Centre Hospitalier Universitaire Hôtel Dieu, Paris, France and Université Paris Descartes, EA 7327, Faculté de Médecine Site Necker, Sorbonne Paris Cité, Paris, France),

Santiago Pérez-Hoyos [GEMES-Haemo, Vall d'Hebrón Institut de Recerca (VHIR), Universitat Autònoma de Barcelona, Barcelona, Spain], Maria Campbell (RCC Copenhagen, Copenhagen, Denmark), Maria Prins [CASCADE, Department of Internal Medicine, Division of Infectious Diseases, Center for Infection and Immunity Amsterdam (CINIMA), Academic Medical Center, Amsterdam, the Netherlands. Dept. Of Infectious Diseases, Public Health Service, Amsterdam, The Netherlands], Geneviève Chêne (RCC Bordeaux, Univ. Bordeaux, ISPED, Centre INSERM U1219-Bordeaux Population Health, Bordeaux, France), Laurence Meyer (SEROCO, INSERM, U1018, Epidemiology of HIV, reproduction, paediatrics, CESP, University Paris-Sud, Le Kremlin-Bicêtre, France, and Department of Public Health and Epidemiology, Bicêtre Hospital, AP-HP, Le Kremlin Bicêtre, France), Maria Dorrucchi (CASCADE, Department of Infectious, Parasitic and Immune-mediated Diseases, Istituto Superiore di Sanità, Rome, Italy), Carlo Torti (MASTER, Unit of Infectious and Tropical Diseases, Department of Medical and Surgical Sciences, University "Magna Graecia", Catanzaro (Italy)) and Rodolphe Thiébaud (Project Lead; Univ. Bordeaux, ISPED, Centre INSERM U1219-Bordeaux Population Health, Bordeaux, France).

References

- Bonnet F, Chene G, Thiebaut R *et al.* Trends and determinants of severe morbidity in HIV-infected patients: the ANRS C03 Aquitaine Cohort, 2000–2004. *HIV Med* 2007; 8: 547–554.
- Nakagawa F, May M, Phillips A. Life expectancy living with HIV: recent estimates and future implications. *Curr Opin Infect Dis* 2013; 26: 17–25.
- Williams I, Churchill D, Anderson J *et al.* British HIV Association guidelines for the treatment of HIV-1-positive adults with antiretroviral therapy 2012 (Updated November 2013. All changed text is cast in yellow highlight.). *HIV Med* 2014; 15 (Suppl 1): 1–6.
- Bofill M, Janossy G, Lee CA *et al.* Laboratory control values for CD4 and CD8 T lymphocytes. Implications for HIV-1 diagnosis. *Clin Exp Immunol* 1992; 88: 243–252.
- Mocroft A, Furrer HJ, Miro JM *et al.* The incidence of AIDS-defining illnesses at a current CD4 count ≥ 200 cells/ μ L in the post-combination antiretroviral therapy era. *Clin Infect Dis* 2013; 57: 1038–1047.
- Young J, Psychogiou M, Meyer L *et al.* CD4 cell count and the risk of AIDS or death in HIV-Infected adults on combination antiretroviral therapy with a suppressed viral load: a longitudinal cohort study from COHERE. *PLoS Med* 2012; 9: e1001194.

- 7 Lewden C, Chene G, Morlat P *et al.* HIV-infected adults with a CD4 cell count greater than 500 cells/mm³ on long-term combination antiretroviral therapy reach same mortality rates as the general population. *J Acquir Immune Defic Syndr* 2007; **46**: 72–77.
- 8 Chêne G, Sterne JA, May M *et al.* Prognostic importance of initial response in HIV-1 infected patients starting potent antiretroviral therapy: analysis of prospective studies. *Lancet* 2003; **362**: 679–686.
- 9 Moore DM, Harris R, Lima V *et al.* Effect of baseline CD4 cell counts on the clinical significance of short-term immunologic response to antiretroviral therapy in individuals with virologic suppression. *J Acquir Immune Defic Syndr* 2009; **52**: 357–363.
- 10 Kjaer J, Ledergerber B. HIV cohort collaborations: proposal for harmonization of data exchange. *Antivir Ther* 2004; **9**: 631–633.
- 11 Geraci M, Bottai M. Quantile regression for longitudinal data using the asymmetric Laplace distribution. *Biostatistics* 2007; **8**: 140–154.
- 12 Charlier I, Paindaveine D, Saracco J. Conditional quantile estimation through optimal quantization. *J Stat Plan Inference* 2015; **156**: 14–30.
- 13 Gilson RJC, Man S-L, Copas A *et al.* Discordant responses on starting highly active antiretroviral therapy: suboptimal CD4 increases despite early viral suppression in the UK Collaborative HIV Cohort (UK CHIC) Study. *HIV Med* 2010; **11**: 152–160.
- 14 Julg B, Poole D, Ghebremichael M *et al.* Factors predicting discordant virological and immunological responses to antiretroviral therapy in HIV-1 clade C infected Zulu/Xhosa in South Africa. *PLoS ONE* 2012; **7**: e31161.
- 15 Le T, Wright EJ, Smith DM *et al.* Enhanced CD4+ T-cell recovery with earlier HIV-1 antiretroviral therapy. *N Engl J Med* 2013; **368**: 218–230.
- 16 Trotta MP, Cozzi-Lepri A, Ammassari A *et al.* Rate of CD4+ cell count increase over periods of viral load suppression: relationship with the number of previous virological failures. *Clin Infect Dis* 2010; **51**: 456–464.
- 17 Abrams D, Levy Y, Losso MH *et al.* Interleukin-2 therapy in patients with HIV infection. *N Engl J Med* 2009; **361**: 1548–1559.
- 18 Levy Y, Sereti I, Tambussi G *et al.* Effects of r-hIL-7 on T cell recovery and thymic output in HIV-infected patients receiving antiretroviral therapy: results of a Phase I/IIa randomized, placebo controlled. Multicenter Study. *Clin Infect Dis* 2012; **55**: 291–300.
- 19 Sterne JA, May M, Costagliola D *et al.* Timing of initiation of antiretroviral therapy in AIDS-free HIV-1-infected patients: a collaborative analysis of 18 HIV cohort studies. *Lancet* 2009; **373**: 1352–1363.
- 20 Abgrall SMIS, May MT, Costagliola D *et al.* Durability of first ART regimen and risk factors for modification, interruption or death in HIV-positive patients starting ART in Europe and North America 2002–2009. *AIDS* (London, England). 2013; **27**: 803–813.
- 21 Autran B, Carcelain G, Li TS *et al.* Positive effects of combined antiretroviral therapy on CD4+ T cell homeostasis and function in advanced HIV disease. *Science* 1997; **277**: 112–116.
- 22 Haynes BF, Markert ML, Sempowski GD, Patel DD, Hale LP. The role of the thymus in immune reconstitution in aging, bone marrow transplantation, and HIV-1 infection. *Annu Rev Immunol* 2000; **18**: 529–560.
- 23 Zeng M, Haase AT, Schacker TW. Lymphoid tissue structure and HIV-1 infection: life or death for T cells. *Trends Immunol* 2012; **33**: 306–314.
- 24 Jameson SC. Maintaining the norm: T-cell homeostasis. *Nat Rev Immunol* 2002; **2**: 547–556.
- 25 Florence E, Lundgren J, Dreezen C *et al.* Factors associated with a reduced CD4 lymphocyte count response to HAART despite full viral suppression in the EuroSIDA study. *HIV Med* 2003; **4**: 255–262.
- 26 Kaufmann GR, Bloch M, Zaunders JJ, Smith D, Cooper DA. Long-term immunological response in HIV-1-infected subjects receiving potent antiretroviral therapy. *AIDS* 2000; **14**: 959–969.
- 27 Le Moing V, Thiébaud R, Chêne G *et al.* Predictors of long-term increase of CD4+ cell count in human immunodeficiency virus-infected patients initiating a protease inhibitor-containing regimen. *J Infect Dis* 2002; **185**: 471–480.
- 28 Kaufmann GR, Perrin L, Pantaleo G *et al.* CD4 T-lymphocyte recovery in individuals with advanced HIV-1 infection receiving potent antiretroviral therapy for 4 years – The Swiss HIV cohort study. *Arch Intern Med* 2003; **163**: 2187–2195.
- 29 Hunt PW, Deeks SG, Rodriguez B *et al.* Continued CD4 cell count increases in HIV-infected adults experiencing 4 years of viral suppression on antiretroviral therapy. *AIDS* 2003; **17**: 1907–1915.
- 30 Smith CJ, Sabin CA, Youle MS *et al.* Factors influencing increases in CD4 cell counts of HIV-positive persons receiving long-term highly active antiretroviral therapy. *J Infect Dis* 2004; **190**: 1860–1868.
- 31 Thiébaud R, Jacqmin-Gadda H, Walker S *et al.* Determinants of response to first HAART regimen in naive patients with an estimated time since HIV seroconversion. *HIV Med* 2006; **7**: 1–9.
- 32 Yotebieng M, Maskew M, Van Rie A. CD4+ gain percentile curves for monitoring response to antiretroviral therapy in HIV-infected adults. *AIDS* 2015; **29**: 1067–1075.

Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

Fig. S1. Percentiles (5th, 10th, 25th, 50th, 75th, 90th, 95th) of CD4+ T cell count over time from cART initiation until 15 months

Table S1. Included vs non-included patients' baseline characteristics

Table S2. CD4 change at 6 months to maintain or accelerate the CD4+ T cell response according